Blue tongue virus in Kenya Insights from Isiolo and progress on molecular typing and virus isolation

Nicholas Svitek
PALE-Blu Blue Tongue Virus Consortium Meeting
Giulianova, Italy
26 September 2019

























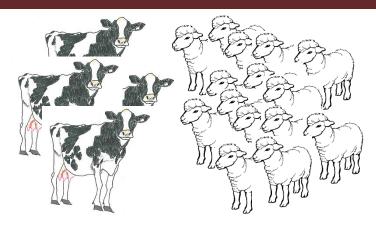
Bluetongue virus (BTV)

Our Involvement in the Project

WP2: Identify and characterize BTV strains circulating in 'source' regions that represent potential threats to Europe



WP2: Work Plan / Study Design



Field sampling during reported outbreaks and in known high prevalence areas

- Serology [ID Screen® Bluetongue Competition (anti-VP7)]
- Molecular epidemiology by PCR (segment 10)/serotype specific qPCR (VP2 gene)
- Sequencing of field samples or
- ➢ Genomic characterization of virus isolates





Isolation of virus on BSR or KC cells by plaque purification





BTV in Kenya

- √ 1978 : 19 different BTV serotypes found in sentinel cattle near Nairobi
 - ➤ Rainfall had no obvious effect on the serum conversion rates.
- ✓ 2013 : In Busia (Western Kenya) 94.2% anti-BTV in cattle (n=455)
 - (single: BTV-7,-15,-16,-19, dual: BTV-1&-12, BTV-3&24, BTV-22&24).
 - Rainfall had no obvious effect on the serum conversion rates.



BTV Outbreak in Kenya-2018



Print ℃lose

Information received on 20/07/2018 from Dr Obadiah Nyaga Njagi, Director of Veterinary Services, Directorate of Veterinary Services, State Department of Livestock, Ministry of Agriculture, Livestock and Fisheries, NAIROBI, Kenya

Summary

Report type	Immediate notification				
Date of start of the event	20/06/2018				
Date of confirmation of the event	25/06/2018				
Report date	20/07/2018				
Date submitted to OIE	20/07/2018				
Reason for notification	Recurrence of a listed disease				
Date of previous occurrence	1998				
Manifestation of disease	Clinical disease				
Causal agent	Bluetongue virus				
Serotype	Not typed				
Nature of diagnosis	Laboratory (basic)				
This event pertains to	a defined zone within the country				
Related reports	Immediate notification (20/07/2018) Follow-up report No. 1 (06/08/2018) Follow-up report No. 2 (15/08/2018) Follow-up report No. 3 (28/08/2018)				

Outbreak 1	Samburu Central, Samburu							
Date of start of the outbreak	20/06/2018							
Outbreak status	Continuing (or date resolved not provided)							
Epidemiological unit	Farm							
Affected animals	Species	Susceptible	Cases	Deaths	Killed and disposed of	Slaughtered		
	Sheep	350	64	0	0	(
Outbreak 2	Gilgil, Nakuru							
Date of start of the outbreak	20/06/2018							
Outbreak status	Continuing (or date resolved not provided)							
Epidemiological unit	Farm							
Affected animals	Species	Susceptible	Cases	Deaths	Killed and disposed of	Slaughtered		
	Sheep	100	10	5	0	(
Outbreak 3	Nakuru, Nakuru							
Date of start of the outbreak	20/06/2018							
Outbreak status	Continuing (or date resolved not provided)							
Epidemiological unit	Farm							
Affected animals	Species	Susceptible	Cases	Deaths	Killed and disposed of	Slaughtered		
	Sheep	150	15	0	0	(



Summary:

of outbreak: 3

Areas: 2 [Samburu and Nakuru]

Species: sheep

Number of cases: 89/600

Deaths: 5

Case fatality rate: **5.62%**



Summary of Years 1 & 2

Year 1

- Getting reagents & protocols
- Getting paper work/approvals
- Training at Pirbright
- Sampling in Machakos
- cELISA with Machakos samples
- Problem with qPCR assay

Year 2

- Finalized cELISA of Machakos samples
- Validated qPCR assay with new probes/primers
- Analyzed blood samples from Machakos by qPCR
- Analyzed Isiolo samples by cELISA & qPCR
- Isolated viruses from the field



BTV in Machakos County – Kapiti Plains





Date of sampling : May 17-18, 2018

Season: towards the end of the long rainy season

Location: Kapiti Plains [ILRI's research station in Machakos]

Coordinates: 1°38'01.2"S 37°08'51.2"E



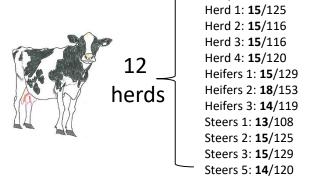


Sampling for BTV in Machakos County

$$n' = \frac{1}{1/n + 1/N}$$

Cattle in Kapiti 2581 – adjusted n = 180Goats in Kapiti 306 – adjusted n = 121Sheep in Kapiti 1326 – adjusted n = 169Camel 37 [~80] in Kapiti – adjusted n = 31 [43]

Total: **513**





3 herds Herd 1: females [milking] Herd 2: females [dry]

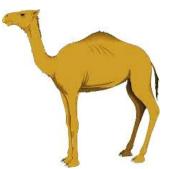
Dairy herd: 18/149

Herd 3: males



3 herds Herd 1: females [older]
Herd 2: females [younger]

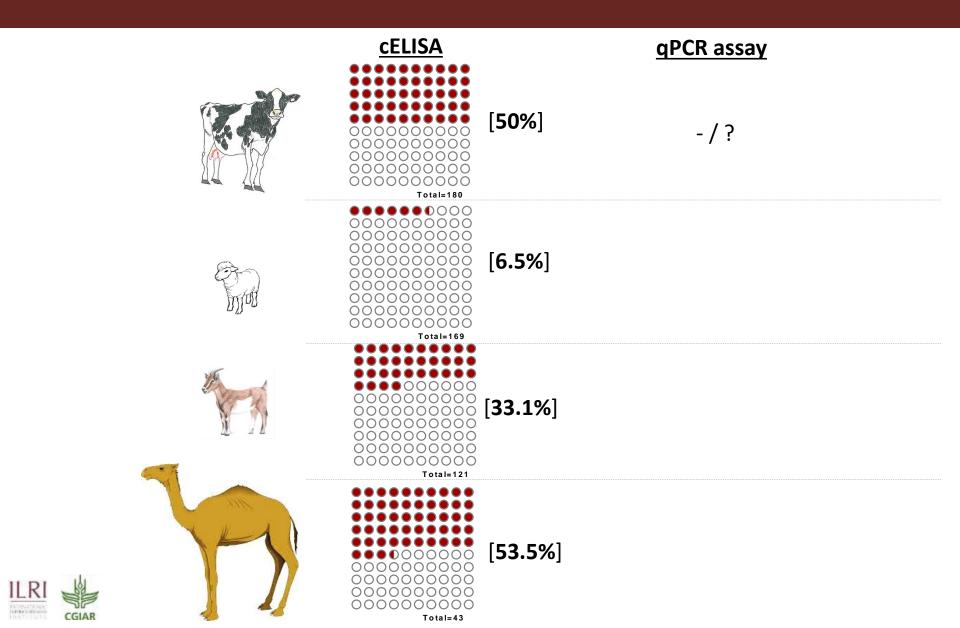
Herd 3: males



1 herd



BTV in Machakos - Results



Validating the BTV qPCR Assay

- BLUVAX vaccine [KEVEVAPI]

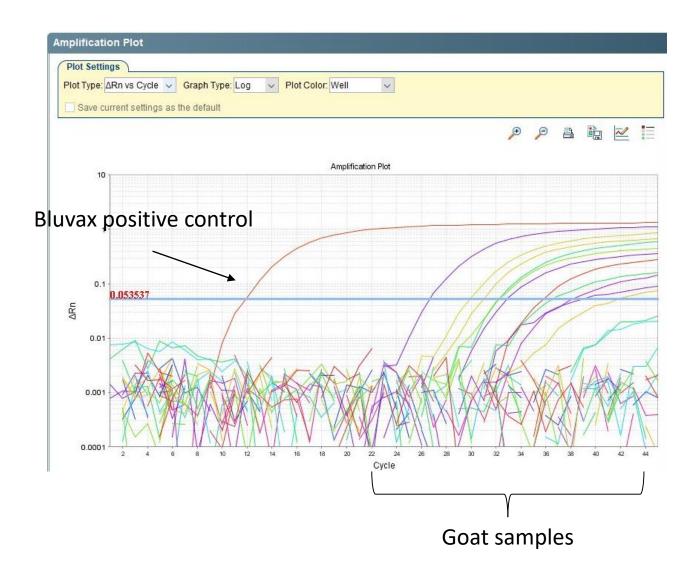


- (BTV 1, 2, 3, 4, 8, 12)
- Panel of BTV-positive samples from Pirbright
 - (2018 BTV Ring Trial panel: BTV 1, 3, 4, 8)
- qPCR with primers/probes for segment 10 (NS3): functional
- qPCR with primers/probes for VP2: functional for serotypes 3, 4, 5, 8, 12



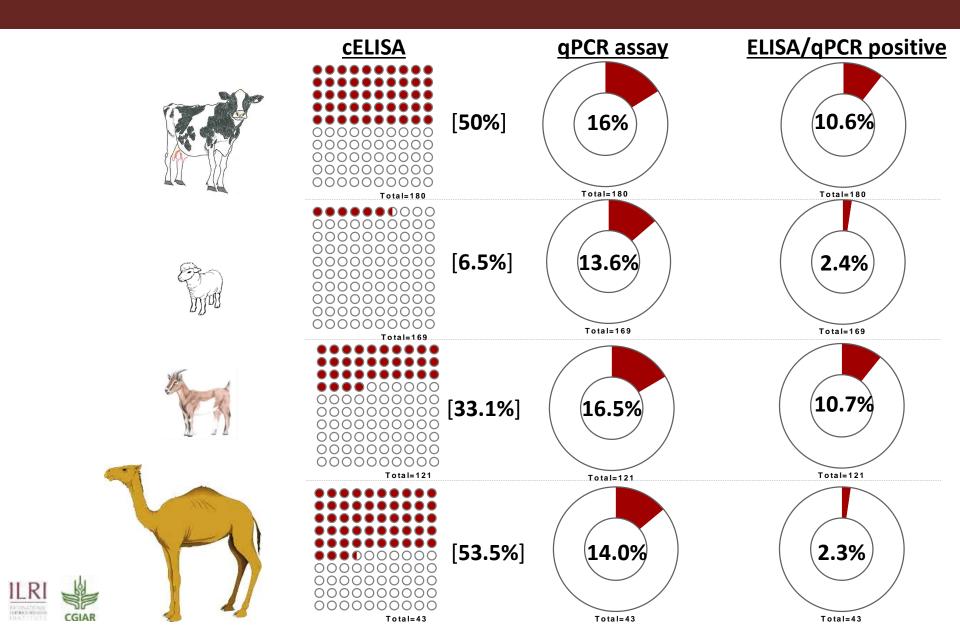
(RNA levels?, probe quality?, new serotype?)

qPCR Assay with Goat Samples from Kapiti

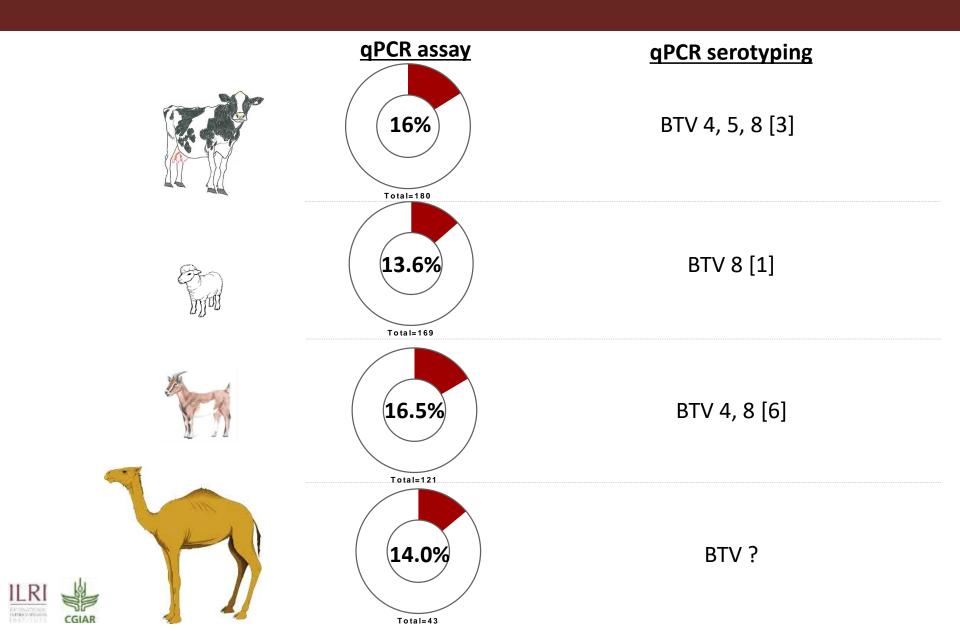




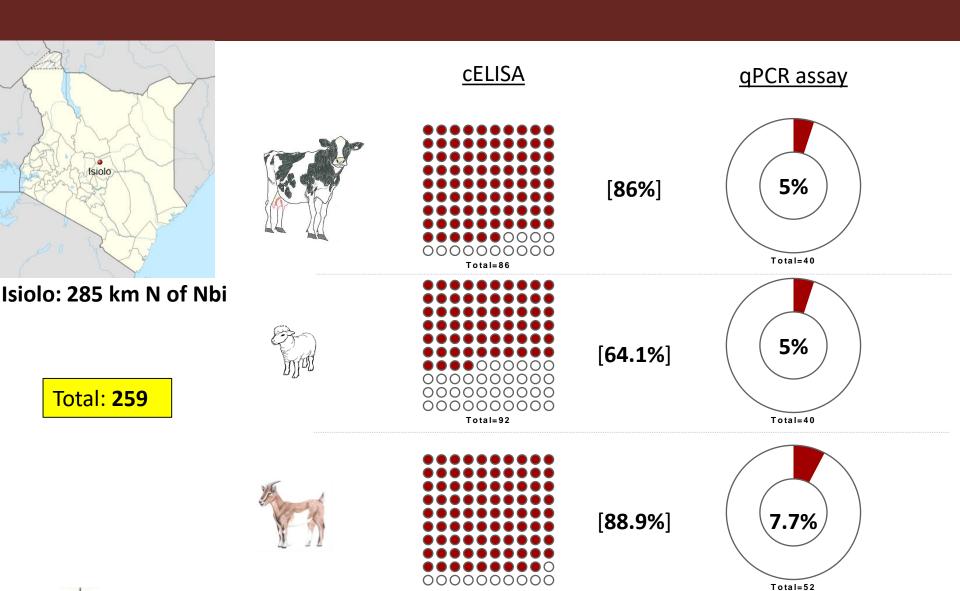
BTV in Machakos - Results



BTV in Machakos – Serotyping Results



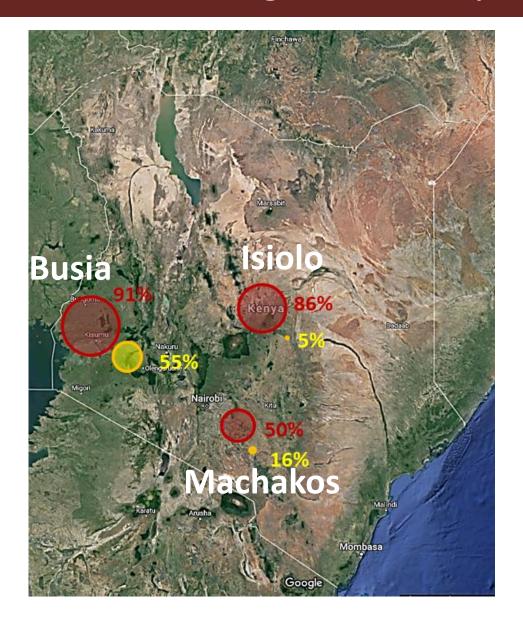
BTV in Isiolo - Results



Total=81



Exposure to BTV in Cattle from Different Regions of Kenya



ELISA

qPCR



Summary – ELISA / qPCR / Serotyping

- Higher prevalence in cattle and camel.
- Higher prevalence of BTV RNA in animals from Busia, followed by Machakos then Isiolo [very low]. (Total of 972 samples tested so far).
- Kenyan sheep have been exposed to BTV [Machakos: 6.5%, Isiolo: 64.1%] without having caused apparent clinical infection [my assumption]?
- Able to type for BTV 3, 4, 5, 8, 12.

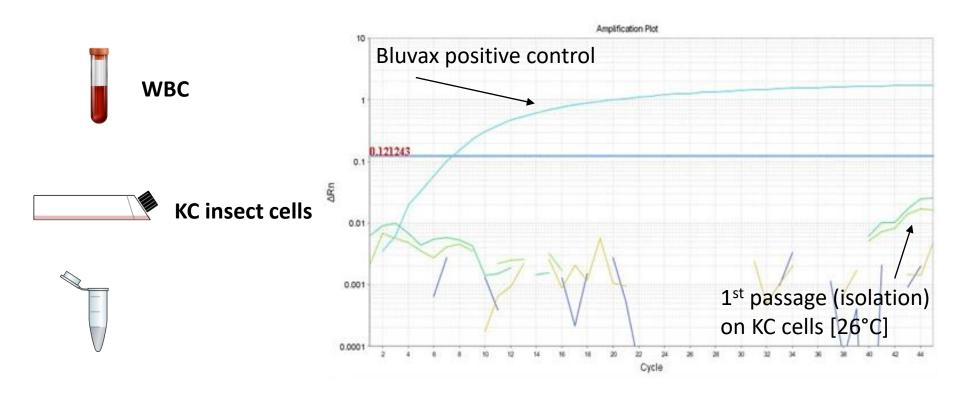




Virus isolation from field samples on KC and BHK cells

BTV Isolates from Kenya – First Trial

2 BTV isolates from sheep



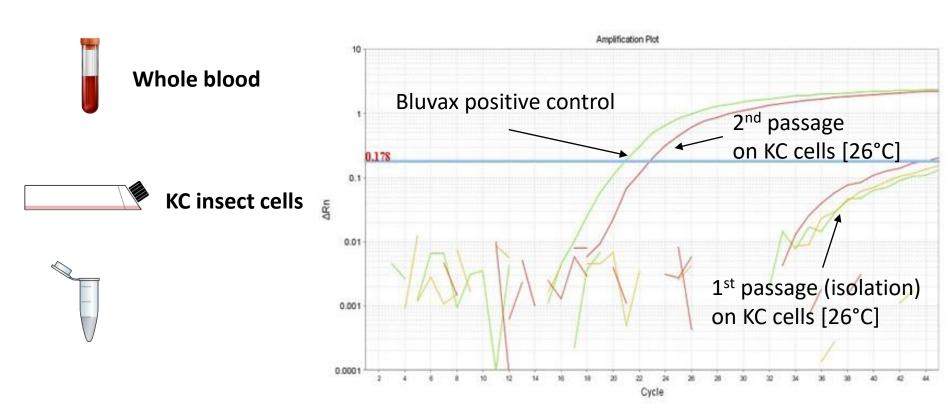
Initial Ct values from blood samples: 28.4, 30.4, 27.5

[Pirbright's cutoff value for virus isolation: Ct of 27 or lower]



BTV Isolates from Kenya – Second Trial

3 BTV isolates from sheep (BTV4, 8, ?)



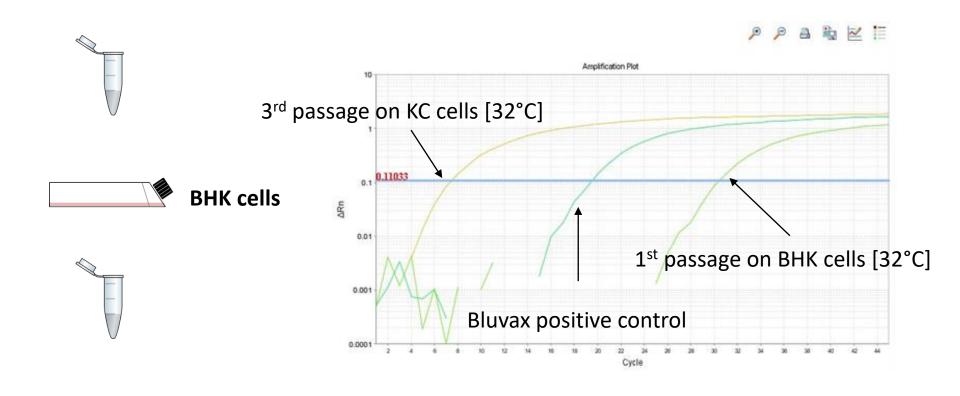


[Pirbright's cutoff value for virus isolation: Ct of 27 or lower]



BTV Isolates from Kenya – Trial on BHK Cells

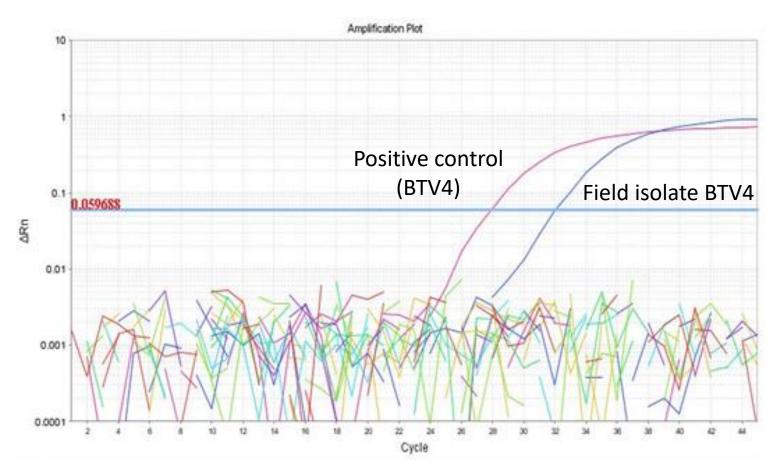
1 BTV isolate from sheep





Serotyping of a field isolate (sheep) – BTV4

Screening with BTV-1 to BTV-26 serotype specific probes [we are only sure that probes for BTV-3,4,5,8,12 work]





Summary – Bluetongue Virus Isolation

- 3 field isolates from sheep.
- 1 BTV 4, 1 BTV 8, one unknown.
- Able to expand virus from one isolate on KC and BHK cells.
- Able to isolate virus from whole WBC as well as from whole blood.



Planning for Next Year & Questions to be Answered

- Obtain samples from Mali & Uganda [ILRI ongoing projects].
- Plan a sampling expedition in Sudan or a country in West Africa [Benin?, Nigeria?].
- Continue qPCR typing and get more field isolates.

- Protocols for plaque-purification of virus isolates?
- Looking at the "BTV RNA conversion rate" instead of Ab conversion rate throughout the year?



Why are Kenyan sheep more resistant to BTV?

Acknowledgments

International Livestock Research Institute



Dr Lucilla Steinaa **Robert Muriuki** Thomas Njoroge

Dr Bernard Bett Josiah Njeru



Dr Carrie Batten Dr John Flannery

NV Reference Lab







Understanding pathogen, livestock, environment interactions involving bluetongue











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