

# Development of diagnostic tools to reduce antimicrobial (mis)use

Novel identified biomarkers and available biobanked samples

Elise Schieck and Sonal Henson Animal and Human Health, ILRI

Scientific seminars ILRI Nairobi, 8 November 2021



### The idea...









## The idea...







ILRI INTERNATIONAL INSTOCK RESEARCH IN STITUTE Today, sick animals are often treated with antimicrobials, regardless of the cause of disease. A diagnostic that differentiates between viral and bacterial infections could potentially reduce this overuse.

## The idea...







To test the possibility of developing an easy-to-use and cheap diagnostic test that can differentiate between bacterial and viral infections.

Bacterial and viral infections typically induce slightly different responses in the hosts

We are testing the possibility of using these host markers to develop a quick and easy-to-use field test.

This may reduce the use of antibiotics to animals infected with viruses.



#### Responses induced by pathogen associated molecular patterns



ILRI INTERNATIONAL UVESTOCK RESEARCH IN STITUTE CGIAR SLU



Discovery group: 240 children 52 definite bacterial infection, 92 definite viral infection, 96 indeterminate infection.

Validation group: 130 children 23 definite bacterial, 28 definite viral, 79 indeterminate infections



Diagnostic Test Accuracy of a 2-Transcript Host RNA Signature JAMA | Preliminary Communication | INNOVATIONS IN HEALTH CARE DELIVERY for Discriminating Bacterial vs Viral Infection in Febrile Children Jethro A. Herberg, PhD; Myrsini Kaforou, PhD; Victoria J. Wright, PhD; Hannah Shailes, BSC; Hariklia Eleftherohorinou, PhD; Clive J. Hoggart, PhD; Miriam Cebev-Lónez. MSc: Michael J. Carter. MRCPCH: Victoria A. Janes. MD: Stuart Gormlev. MRes: Chisato Shimizu. MD: Adriana H. Tremoulet. Jethro A. Herberg, PhD; Myrsini Kaforou, PhD; Victoria J. Wright, PhD; Hannah Shailes, BSc; Hariklia Eleftherohorinou, PhD; Clive J. Hoggart, PhD; Miriam Cebey-López, MSc; Michael J. Carter, MRCPCH; Victoria A. Janes, MD; Stuart Gormley, MRes; Chisato Shimizu, MD; Adriana H. Tremoulet, MD; Anouk M. Barendreet. BSc: Antonio Salas. PhD: John Kanegave. MD: Andrew J. Pollard. PhD: Saul N. Faust. PhD: Saniav Patel. FRCPCH: Miriam Cebey-López, MSc; Michael J. Carter, MRCPCH; Victoria A. Janes, MD; Stuart Gormley, MRes; Chisato Shimizu, MD; Adriana H. Tré Anouk M. Barendregt, BSc; Antonio Salas, PhD; John Kanegaye, MD; Andrew J. Pollard, PhD; Saul N. Faust, PhD; Sanjay Patel, Frederic Taco Kujiners, PhD; Federico Martinón-Torres, PhD; Jane C. Rume, MD; Lachlan, LM. Coin, PhD; Michael Levin, ERCPCH, for the Interna-Anouk M. Barendregt, BSc; Antonio Salas, PhD; John Kanegaye, MD; Andrew J. Pollard, PhD; Saul N. Faust, PhD; Sanjay Patel, FRCPCH; Taco Kuijpers, PhD; Federico Martinón-Torres, PhD; Jane C. Burns, MD; Lachlan J. M. Coin, PhD; Michael Levin, FRCPCH; for the IRIS Consortium

- Identified 38 transcripts differentially expressed
- Narrowed down to 2-transcript ٠ ration to discriminate bacterial vs. viral infections



#### Setting up...

- Homologues of top candidates identified in pigs
- Candidate reference genes identified
- Primers for qPCR designed
- qPCR set up (normalized to reference genes)

| Category               | Gene         |  |
|------------------------|--------------|--|
| Putative viral markers | IFNα         |  |
|                        | IFNβ         |  |
|                        | IFITM3       |  |
|                        | STING        |  |
|                        | IFI44L       |  |
|                        | IFIT3        |  |
|                        | MxA          |  |
|                        | RSAD2        |  |
| Putative Bacterial     | FAM89A       |  |
| markers                | S100PBP      |  |
|                        | SLPI         |  |
|                        | UPB1         |  |
| Pro-inflam-matory      | IL-1β        |  |
| cytokines              | IL-6         |  |
|                        | IL-8         |  |
|                        | $TNF-\alpha$ |  |



#### **Test material**

- Pig PBMCs stimulated with agonists
- Pig PBMCs stimulated with split influenza virus or inactivated Actinobacillus pleuropneumoniae

| Category               | Gene         |
|------------------------|--------------|
| Putative viral markers | IFNα         |
|                        | IFNβ         |
|                        | IFITM3       |
|                        | STING        |
|                        | IFI44L       |
|                        | IFIT3        |
|                        | MxA          |
|                        | RSAD2        |
| Putative Bacterial     | FAM89A       |
| markers                | S100PBP      |
|                        | SLPI         |
|                        | UPB1         |
| Pro-inflam-matory      | <i>IL-1β</i> |
| cytokines              | IL-6         |
|                        | IL-8         |
|                        | TNF-α        |



| Category                       | Gene        |                    | Viral mimics      |                      | Bacterial mimics      |               |                | Inactivated microbes |                   |
|--------------------------------|-------------|--------------------|-------------------|----------------------|-----------------------|---------------|----------------|----------------------|-------------------|
|                                |             | ODN 2216<br>(TLR9) | R848 (TLR7/<br>8) | poly (I:C)<br>(TLR3) | Pam3CSK4 (TLR2/<br>1) | LPS<br>(TLR4) | FLiC<br>(TLR5) | Split<br>Influenza   | A. pleuro<br>(HI) |
|                                | IFNα        | 3.4                | 0                 | 1.0                  | 0.2                   | 0.3           | 0.3            | 1.7                  | 0.4               |
|                                | IFNβ        | 409.2              | 1.8               | 1992.0               | 3.0                   | 0.5           | 1.7            | 70.9                 | 1.4               |
|                                | IFITM3      | 19.9               | 11.9              | 5.7                  | 2.7                   | 0.9           | 1.3            | 22.5                 | 0.7               |
|                                | STING       | 0.6                | 0.2               | 0.6                  | 0.5                   | 0.5           | 0.7            | 1.3                  | 0.4               |
|                                | IFI44L      | 7.5                | 3.4               | 3.3                  | 1.4                   | 0.7           | 1.1            | 8.0                  | 0.5               |
|                                | IFIT3       | 41.7               | 12.4              | 7.0                  | 1.5                   | 46.2          | 0.7            | 95.9                 | 0.2               |
| -                              | MxA         | 72.6               | 39.5              | 17.7                 | 7.9                   | 0.5           | 1.0            | 63.0                 | 0.7               |
|                                | RSAD2       | 206.7              | 73.3              | 30.7                 | 7.6                   | 0.8           | 1.5            | 113.5                | 0,5               |
| Putative Bacterial             | FAM89A      | 0.6                | 0.1               | 0.5                  | 0.5                   | 0.8           | 0.8            | 0.6                  | 0.6               |
| markers                        | S100PBP     | 0.6                | 0.2               | 0.7                  | 0.6                   | 0.5           | 0.6            | 0.7                  | 0.4               |
|                                | SLPI        | 72.6               | 5.3               | 28.1                 | 5.0                   | 0.2           | 2.0            | 233.8                | 0.3               |
|                                | UPB1        | 25.5               | 78.5              | 5.9                  | 66.5                  | 38.8          | 20.1           | 2.2                  | 30.8              |
| Pro-inflam-matory<br>cytokines | IL-1β       | 1.0                | 1.8               | 10.3                 | 67.2                  | 40.7          | 30.0           | 0.7                  | 17.7              |
|                                | IL-6        | 20.7               | 96.7              | 19.7                 | 72.3                  | 51.7          | 24.5           | 3.7                  | 12.2              |
|                                | IL-8        | 1.9                | 9.0               | 44.0                 | 119.8                 | 66.6          | 32.0           | 0.3                  | 24.3              |
|                                | TNF-α       | 2.6                | 0.9               | 2.3                  | 1.9                   | 2.4           | 2.5            | 2.1                  | 1.6               |
| ≤ 0.0625 > 0.0                 | 625 - 0.125 | > 0.125 - 0        | ).25 > 0.25       | 5 - 0.5 > 0.         | 5 - < 2 2 to < 4      | 4 4 to        | 0 < 8          | 8 to < 16            | ≥16               |

#### Hjertner et al., PLOS ONE 2021

#### Testing on blood from Actinobacillus pleuropneumoniae infected pigs



**CGIAR** 

INSTITUTE





## Phase 2

Goal: To expand the list of candidate genes to test On more samples

Method: Identify genes from published transcriptome studies in pigs.



## Transcriptome studies included in analysis

| Viral-infection  | Bacterial-infection  | Virus-Bacterial co-infection   |
|--|--|--|
| Cruz-Pulido et al. (2021)<br>- Comparative Transcriptome Profiling of<br>Human and Pig Intestinal Epithelial Cells<br>after Porcine Deltacoronavirus (PDCov)<br>Infection.<br>- RNASeq   | Kamminga et al. (2020)<br>- Combined Transcriptome Sequencing of<br><u>Mycoplasma hyopneumoniae</u> and Infected Pig<br>Lung Tissue Reveals Up-Regulation of Bacterial<br>F1-Like ATPase and Down-Regulation of the<br>P102 Cilium Adhesin in vivo<br>- RNASeq | <ul> <li>Dang et al. (2014)</li> <li>Transcriptional approach to study porcine tracheal epithelial cells individually or dually infected with <u>swine influenza virus</u> and <u>S. suis</u></li> <li>Microarray</li> </ul>                   |
| <ul> <li>Miller et al. (2020)</li> <li>Comparison of the transcriptome response within the swine tracheobronchial lymph node following infection with <u>PRRSV, PCV-2</u> <u>or IAV-S</u>.</li> <li>Digital Gene Expression Tag Profiling (DGETP)</li> </ul> | Ni et al. (2019)<br>- RNA-seq transcriptome profiling of porcine<br>lung from two pig breeds in response to<br><u>Mycoplasma hyopneumoniae</u> infection.<br>- RNASeq  | Lin et al. (2015)<br>Investigation of Pathogenesis of <u>H1N1</u><br><u>Influenza Virus</u> and <u>Swine Streptococcus</u><br><u>suis</u> Serotype 2 Co-Infection in Pigs by<br><u>Microarray</u> Analysis                                     |
| Liang et al. (2017) – PRRSV-infected PAMs<br>Dong et al. (2021) – PRRSV-infected tonsils<br>Hu et al. (2020) - PEDV-infected IPEC  | Yan et al. (2019)<br>- Histological and comparative<br>transcriptome analyses provide insights into<br>small intestine health in diarrheal piglets<br>after infection with <u>Clostridium perfringens</u><br>type C.<br>- RNASeq                               | Auray et al. (2016)<br>- Transcriptional Analysis of <u>PRRSV</u> -<br>Infected Porcine Dendritic Cell Response<br>to <u>Streptococcus suis</u> Infection Reveals<br>Up-Regulation of Inflammatory-Related<br>Genes Expression<br>- Microarray |

GIAR

## Pathways upregulated in the studies







- 1. Clean and merge data from all selected studies
  - There are many gaps in the data, e.g there are no genes where we have data from all pathogen infections.
  - Different methods used for measuring gene expression
- 2. Identify genes with similar expression profiles by clustering
  - UR in viral-infected but DR in bacterial-infected
  - UR in bacterial-infected but DR in viral-infected



## Final datasets chosen for comparison

#### 1. Microarray datasets

- 1. Dang et al., 2014
  - Infection with <u>H1N1</u>, <u>Streptococcus suis</u>, and co-infection with both in porcine tracheal epithelial cells
- 2. Lin et al., 2015
  - Pigs infected with <u>H1N1</u>, <u>S. suis</u> and co-infection with both.
  - Expression in lung tissue.
- 3. Auray et al., 2016
  - Infection of dendritic cells and monocytes with <u>S. suis</u> and <u>PRRSV</u>.





## Genes UR by bacterial infection in at least 2 studies

| Function                                     | SYMBOL | Gene description  |
|--|--------|---|
| Biological and metabolic processes           | HK2    | Hexokinase 2  |
|  | PLAT   | plasminogen activator, tissue                                       |
|  | SLC2A1 | Solute carrier family 2 (facilitated glucose transporter), member 1 |
| Cytokines, chemokines, and related receptors | CSF2   | colony stimulating factor 2   |
|  | SPP1   | Secreted phosphoprotein 1   |
| Lipid metabolism                             | LDLR   | Low density lipoprotein receptor                                    |
| Transcriptional and translational regulation | AGO2   | argonaute RISC catalytic component 2                                |



## Genes UR by viral infection in at least 2 studies

| Function                                   | SYMBOL  | Gene description  |  |  |
|--|---------|---|--|--|
| Biological & metabolic processes           | USP18   | ubiquitin specific peptidase 18                             |  |  |
|  | ZBP1    | Z-DNA binding protein 1                                     |  |  |
|  | BCR     | BCR activator of RhoGEF and GTPase                          |  |  |
| Cytokine signalling                        | EIF2AK2 | eukaryotic translation initiation factor 2 alpha kinase 2   |  |  |
|  | TRIM21  | tripartite motif containing 21                              |  |  |
| Cytokines, chemokines, & related receptors | CCL4    | C-C motif chemokine ligand 4                                |  |  |
|  | TNFSF10 | TNF superfamily member 10                                   |  |  |
|  | IFNB1   | interferon beta 1   |  |  |
| Cytoskeleton/actin rearrangement           | TMOD4   | tropomodulin 4  |  |  |
|  | RSAD2   | radical S-adenosyl methionine domain containing 2           |  |  |
|  | IFIT3   | interferon-induced protein with tetratricopeptide repeats 3 |  |  |
|  | IFIT1   | interferon-induced protein with tetratricopeptide repeats 1 |  |  |
|  | MX1     | MX dynamin like GTPase 1                                    |  |  |
| Defence response                           | MX2     | myxovirus (influenza virus) resistance 2 (mouse)            |  |  |
|  | DDX58   | DExD/H-box helicase 58                                      |  |  |
|  | OAS2    | 2'-5'-oligoadenylate synthetase 2                           |  |  |
|  | GBP1    | guanylate binding protein 1, interferon-inducible           |  |  |
|  | IFIH1   | interferon induced with helicase C domain 1                 |  |  |
|  | PARP12  | poly(ADP-ribose) polymerase family member 12                |  |  |
| Transcriptional & translational regulation | PARP14  | poly(ADP-ribose) polymerase family member 14                |  |  |

#### Short term:

- Test more biomarkers
- Collect more samples

#### Long term:

- Expand to other livestock
- Transfer to penside format

#### Samples collected for future use

| disease | species | healthy | disease | sample type<br>(full blood) |
|---------|---------|---------|---------|-----------------------------|
| СВРР    | cattle  | 15      | 15      | paxgene                     |
| ССРР    | goats   | 40      | 28      | paxgene                     |
| СВРР    | cattle  | 31      | 9       | RNAlater                    |
| AFS     | pigs    | 17      | 5       | RNAlater                    |
| BRSV    | cattle  | 54      |         | RNAlater                    |



### Israel's MeMed gets FDA approval for 'breakthrough' infection test

Purpose of test is to tell physicians whether body is waging war on bacteria or virus, and make decisions about whether to treat with antibiotics

By RICKY BEN-DAVID ~ 20 September 2021, 5:14 pm |





MeMed Diagnostics founders Dr. Kfir Oved and Dr. Eran Eden. (Courtesy)



#### Acknowledgements

**ILRI** Team

SLU Team

SVA

Elise Schieck Sonal Henson Benjamin Nzau Caroline Fossum Bernt Hjertner Claudia Lützenschwab Ulf Magnusson

Marie Sjölund









The International Livestock Research Institute (ILRI) is a non-profit institution helping people in low- and middle-income countries to improve their lives, livelihoods and lands through the animals that remain the backbone of small-scale agriculture and enterprise across the developing world. ILRI belongs to CGIAR, a global research-for-development partnership working for a food-secure future. ILRI's funders, through the <u>CGIAR Trust Fund</u>, and its many partners make ILRI's work possible and its mission a reality. Australian animal scientist and Nobel Laureate Peter Doherty serves as ILRI's patron. You are free to use and share this material under the Creative Commons Attribution 4.0 International Licence @①.

better lives through livestock

ilri.org